The Site Frequency Spectrum

Hancock February 8, 2024

Ten independently generated gene genealogies (2N=20)

Genealogies generated under constant population size, random mating, no selection

Wide variation simply due to probabilistic nature of the timing of coalescence events

Coalescent Models, Wakeley, from Lohmueller and Nielsen

The site frequency spectrum (SFS)

The SFS is a histogram of allele counts



Note that: in different contexts, axes may be expressed as counts or as proportions (or probabilities)

Coalescent Models, Wakeley, from Lohmueller and Nielsen

A site's position in the spectrum depends on its position in the gene tree



Mutations A and C are singletons; B is a doubleton

Most recent interval: singletons only

2nd most recent: singletons and doubletons

3rd most recent: singletons, doubletons, and tripletons

A tree with 2 leaves has only singletons



We expect $4Nu = \theta$ mutations, all singletons.

Number of branches x $L \times u = 2 \times 2N \times u = 4Nu$

With 3 leaves, there are the same number of singletons but half as many doubletons



A.R. slides

The expected spectrum in a population of constant size

Sample	Exp	Expected spectrum										
size	(sin	(singletons, doubletons,)										
2	θ											
3	θ ,	$\theta/2$										
4	θ ,	$\theta/2,$	$\theta/3$									
5	θ ,	$\theta/2,$	$\theta/3,$	$\theta/4$								
Etcetera												

Note that as we increase the sample size, the expected number of mutants in each category stays the same

A neutral site frequency spectrum



So, a neutral (unfolded) SFS looks something like this, where the number of doubletons is about half the number of singletons, and the number of tripletons is about 1/3 the number of singletons, ...

Coalescent Models, Wakeley, from Lohmueller and Nielsen

A coalescent genealogy with variant sites





How many singletons?

How many doubletons?

Coalescent Models, Wakeley, from Lohmueller and Nielsen

Relationship between a genealogy, sequence data, and the SFS

- 8 chromosomes ("genes") are sampled from the population
- This could be from 4 diploid individuals in a randomly mating population
- This is a region with no history of recombination



Relationship between a genealogy, sequence data, and the SFS



Convert an unfolded SFS to a folded SFS



In the folded SFS, we only use information about which is the minor and which is the major allele, so the categories for the extremes are grouped (i.e., 1&9, 2&8, 3&7, 4&6 become 1, 2, 3, and 4)

How to calculate *S* from the folded SFS?



How to calculate *S* from the folded SFS?



To get S, just add up the counts from each category

$$S = 6 + 3 + 4 + 2 + 1$$

= 16

How to calculate (per sequence*) $\hat{\theta}_{S}$ from the folded SFS?





*recall: to calculate $\hat{\theta}_S$ per nucleotide, you would need to know the total number of assayed sites

How to calculate π from the folded SFS

Recall from Gene genealogies lecture:

	00000	00001								
	12345	67890								
S1	AAACT	GTCAT								
S2		A								
S 3		AC								
S4	G	Α								
S5	G	A								
S6	G	A								
	^	^								
	1	1								
	1		Contributes	1	X	5	=	5	pairwise	diffs
			Contributes	3	Х	3	=	9	pairwise	diffs

...Sum these up and divide by the number of pairwise comparisons

See gene genealogies lecture notes sec 1.3 for more info

How to calculate (per sequence*) $\hat{\theta}_{\pi}$ from the folded SFS?



$$\hat{\theta}_{\pi} = \pi$$

$$=\frac{6(1 \times 9) + 3(2 \times 8) + 4(3 \times 7) + 2(4 \times 6) + 1(5 \times 5)}{45}$$

$$= \frac{54 + 48 + 84 + 48 + 25}{45} = 5.76$$

*recall: to calculate $\hat{\theta}_{\pi}$ per nucleotide, you would need to know the total number of assayed sites

Site frequency spectra under different population history models



Coalescent Models, Wakeley, from Lohmueller and Nielsen

Cartoon depictions of genealogies from the four different population history models



Coalescent Models, Wakeley, from Lohmueller and Nielsen

Standard neutral population



This SFS fits has roughly the expected distribution of frequencies (θ /*i*) for singletons, doubletons, tripletons, etc: θ , θ /2, θ /3, ...

Recent 100-fold population growth



An excess of low frequency variants in the SFS due to rapid population expansion

Two isolated populations



Population subdivision results in an excess of intermediate allele frequencies in the SFS because alleles often coalesce farther back in time

One deme in a subdivided population

A subdivided population in which migration can occur among five local populations



There are multiple ways to measure nucleotide diversity (θ)

The most popular estimates are:

- Waterson's θ_W , which is also called θ_S (based on S, the number of segregating sites)
- Tajima's θ_{π} (based on π , the number of pairwise differences)

There are multiple ways to measure nucleotide diversity (θ)

- Waterson's θ (based on S, the number of segregating sites)

$$\widehat{\theta}_S = \frac{S}{\sum_{i=1}^{K-1} \frac{1}{i}}$$

- $\hat{\theta}_{\pi}$ (based the number of pairwise differences)

$$\hat{\theta}_{\pi} = \frac{\sum_{i < j} \pi(i, j)}{\binom{n}{2}}$$

 $\pi(i, j)$ is the number of differences between two sequences n is the number of sequences in the sample

Comparing estimates of θ provides insights into the history of a population or locus

- Different theta estimates summarize different aspects of the site frequency spectrum (and different patterns of variation on the genealogy)
- By comparing these different estimates of theta, we can compare these different aspects of the site frequency spectrum (and the genealogy)
- There are several statistics that have been created to provide a way to summarize such comparisons. The most popular is called *Tajima's D*
- Events that occurred in the history of a *population* create *genome-wide* deviations from the expectation under random-mating
- In comparison, *selective events affect single loci* and create deviations in the statistics *at a particular locus* relative to the rest of the genome

Tajima's D statistic

Compares estimates of θ based on the number of segregating sites (S) and π (the number of pairwise differences) in the sample

$$D = \frac{\widehat{\theta}_{\pi} - \widehat{\theta}_{S}}{\sqrt{\widehat{\mathbf{Var}}[\widehat{\theta}_{\pi} - \widehat{\theta}_{S}]}}$$



Genome-wide patterns of Tajima's D are impacted by population history



Can get this genome-wide from population subdivision or a bottleneck Can get this genome-wide from recent population growth



 $\theta_{\pi} < \theta_{W}$

D negative

Single-locus patterns of Tajima's D are impacted by selection



Can get this in a single region from balancing selection

Can get this in a single region from a selective sweep

 $\theta_{\pi} < \theta_{W}$ D negative



Real examples: human mitochondrial DNA



Represents expected value
 Bars represent observed values

In mtDNA, there is an excess of singletons relative to expected

from Gene genealogies lecture notes

Real examples: human Y chromosome

$$\begin{array}{c}
 Y^{b,c} \\
 K = 718 \\
 S = 20 \\
 \end{array}$$

Represents expected value
 Bars represent observed values

On the Y chromosome, there is an excess of singletons relative to expected

from Gene genealogies lecture notes

Real examples: human beta globin



Represents expected value
 Bars represent observed values

At the Beta Globin locus, there is an excess of intermediate frequency alleles relative to expected

But why?

Some polymorphisms in β -globin are thought to protect against malaria but also to lead to sickle-cell anaemia and thalassemia, they are under balancing selection

from Gene genealogies lecture notes

Comparing two populations using the joint site frequency spectrum (JSFS)

The joint site frequency spectrum includes information about sharing of alleles and their frequencies within and between populations

Counts of alleles in each frequency category in population 2

> Counts of alleles in each frequency category in population 1

Alleles that segregate in population 2 but not in population 1

Counts of alleles in each frequency category in population 2 Derived

allele

b

ibsent

Ы

dod

Derived allele

fixed in

dod

Р

Counts of alleles in each frequency category in population 1



frequency category in population 1

Alleles that segregate in population 1 but not in population 2

Counts of alleles in each frequency category in population 2

Counts of alleles that segregate in both populations grouped by their joint frequencies

Counts of alleles in each frequency category in population 1





Simulated a simple split, immediately after the split

The impacts of demographic history on the JSFS



An example: human populations from Africa (YRI), Europe (CEU), and China (CHB)



Fitting a demographic model to the JSFS



Gutenkunst et al., 2009

An example: Arabidopsis thaliana from Cape Verde



Fulgione, Neto et al., Nature Communications 2022 https://www.nature.com/articles/s41467-022-28800-z

Cvi-0: an enigmatic Arabidopsis accession



A single Arabidopsis plant (Cvi-0) was collected >35 years ago in the Cape Verde Islands, but it was not clear how it got there

History of the Cape Verde Islands



- Colonized by Portuguese in 1460
- Current flora is a mix of endemics and species introduced since colonization
- Main inputs of endemic flora derive from Africa and the Canary Islands



Arabidopsis in Cape Verde



Arabidopsis is present on two islands in Cape Verde





Madeiran colonization: Fulgione et al., MBE 2018 African Arabidopsis: Durvasula, Fulgione et al., PNAS 2017 1001 Genomes: Alonso Blanco et al., Cell 2016

CVI populations represent a single migration from North Africa

- CVI nested within Moroccan clade
- Divergence to Morocco is shared between islands
- Diversity in CVI is low
 - Morocco $\theta_{\rm W}$ = 5.56 x10⁻³
 - \circ Santo Antao θ_{W} = 7.59x10⁻⁵
 - Fogo $\theta_{W} = 8.93 \times 10^{-5}$



Fulgione, Neto et al., Nature Communications 2022 https://www.nature.com/articles/s41467-022-28800-z

CVI lineages are phylogenetically distinct



Fulgione, Neto et al., Nature Communications 2022 https://www.nature.com/articles/s41467-022-28800-z

Split time based on mean pairwise divergence across samples



Inferring population history from sequence data

ARG-based methods use information from across the genome to infer coalescence times between chromosomes



Split time based on the distributions of coalescence times across the genome



Given the rapid population expansion, would you expect Tajima's D to be positive or negative?



Ahmed Elfarargi

Fulgione*, Neto* et al., Nat. Comm., 2022

Tajima's D across Arabidopsis populations

Cape Verde has a very negative Tajima's D



Overall picture: CVI islands were colonized approximately 5 kya through a natural event



Fulgione*, Neto* et al., Nat. Comm., 2022

SFS Summary

- The site frequency spectrum (SFS) is a histogram of allele frequencies within a sample. It summarizes the count of alleles at each frequency in the sample.
- In a randomly mating population, under neutrality and constant population size, θ/i is the expected number of sites at which the derived allele is present in i copies. Note that this does not depend on sample size
- Genome-wide departures from this model imply something about the history of the population as a whole
- Locus-specific departures from this model imply selection specific to that locus
- Tajima's D is a statistic that allows you to compare different aspects of the frequency spectrum (different estimates of θ) to determine whether there is a departure from the model
- The Joint Site Frequency Spectrum (JSFS) summarizes the degree of sharing between two populations. It can be used to infer historical split times and migration rates.